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Clinical-functional and immune-microbiological features of the oral cavity in children and adolescents with chronic forms of gingivitis

DSobirov Alijon Axmadovich1*, DGafforov Sunnatullo Amrulloevich2, DNurmatova Nodira

Tukhtakhujaevna³, ^DDjambilov Ravshan Sotvoldievich⁴, ^DYakubova Farida Xaldarovna⁵

DRadjabov Nodirxon Matlubovich⁶, DGafforova Sabina Sunnatulloyevna⁷, DIbragimov Faxriddin Nusratovich⁸

^{12,3,6}Center for the Development of Professional Qualifications of Medical Workers under the Ministry of Health of the Republic of Uzbekistan, Parkent Street 51, Mirzo-Ulugbek District, Tashkent, Uzbekistan; alijonsobirov204@gmail.com (S.A.A.) gafforovsunnatullos@gmail.com (G.S.A.) nurmatovanodira@gmail.com (N.N.T.). n.radjabovinc@gmail.com (R.N.M.). ⁴Andijan State Medical Institute, Andijan Region, Andijan City, Y. Otabekov Street, Andijan, Uzbekistan, jambilovr@gmail.com (D.R.S.).

⁵Tashkent Pediatric Medical Institute, Bogishamol St 223, Tashkent, Uzbekistan; dr.farida1973@gmail.com (Y.F.X.).

⁷Tashkent State Dental Institute, Tashkent city, Yashnobod district, Makhtumkuli street, house 103, Uzbekistan; gaffarovasabinaa@gmail.com (G.S.S.).

⁸Tashkent State University of oriental studies, Tashkent city, Amir Temur street 20, Uzbekistan, Postal Code: 100060; fahriddin0414@mail.ru (I.F.N.).

Abstract: It is known that inflammatory periodontal diseases (IPD) rank second among dental diseases and are widespread across all population categories regardless of age, place of residence, or gender. It is also known that the transition from gingivitis to periodontitis begins with the appearance of a specific group of oral cavity pathogens, among which P. gingivalis, Actinobacillus actinomycetemcomitans, Treponema denticola, and others play a significant role. Local causes of IPD include poor oral hygiene, improper techniques and use of basic and additional oral care products, frequent consumption of soft foods, and an excess of easily fermentable carbohydrates. Currently, the prevalence of IPD has sharply increased and continues to rise, with a growing trend among younger individuals. Based on the above, this study examines the clinical-functional and immunomicrobiological state of the oral cavity, including the gingival sulcus, in children and adolescents suffering from catarrhal and hypertrophic gingivitis. A total of 425 children and adolescents were selected for the study, including those with chronic catarrhal gingivitis (n=195, CCG) and chronic hypertrophic gingivitis (n=165, CHG), as well as practically healthy children (n=65, control group - CG). Based on the established diagnosis, the clinical-functional and immunomicrobiological state of the oral cavity, including the gingival sulcus, was examined in children and adolescents with various forms of gingivitis. It was found that children and adolescents with CCG and CHG exhibited poor oral hygiene, which worsened with age. Severe inflammatory processes in the gums were observed in all children and adolescents with chronic gingivitis, particularly catarrhal gingivitis. Additionally, local circulatory disorders in the gums were noted, accompanied by increased vascular tone, decreased peripheral resistance to blood flow, reduced vascular elasticity, and impaired blood rheology. Furthermore, β -hemolytic streptococci of group A were identified as the predominant microorganisms in the gingival sulcus and oral cavity, along with a significant increase in the proportion of Candida spp., particularly Candida albicans. Immune status disorders were also detected, characterized by increased IgG, IgA, and IgM concentrations in the blood. A correlation was observed between the quantitative indicators of microorganisms in oral fluid and gingival sulcus contents in patients with CCG and CHG. Based on the OHI-S and PLI indices, as well as caries indicators of primary and permanent teeth in patients with CCG, poor oral hygiene was evident (p

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^{*} Correspondence: alijonsobirov204@gmail.com

<0.0001). The results of various functional and clinical assessments suggest that patients with CCG and CHG exhibit local circulatory disorders in chronic disease progression. These disorders are accompanied by increased vascular tone, while β -hemolytic streptococci of group A and Candida albicans frequently dominate in the gingival sulcus and oral cavity. Additionally, a correlation was found between local immune status, particularly among children aged 10–13 years.

Keywords: Chronic catarrhal gingivitis, Chronic hypertrophic gingivitis, Dentistry, immunology, Functional research methods of the oral cavity, Inflammatory periodontal diseases, Oral hygiene, Oral microbiology.

1. Introduction

Relevance. It is known that inflammatory periodontal tissue diseases (IPTD) rank second among dental diseases and are widespread across all population categories, regardless of age, place of residence, or gender [1-3]. It is also established that the primary microbial composition of purulent-inflammatory processes in the maxillofacial region consists of both facultative anaerobic microorganisms and obligate aerobes [4-6]. More recent studies demonstrate that the transition from gingivitis to periodontitis begins with the emergence of a specific group of oral pathogens (OP), among which P. gingivalis, *Actinobacillus actinomycetemcomitans, Treponema denticola*, and others play a significant role [7-9].

Studies have proven that local causes of inflammatory periodontal tissue diseases (IPTD) include: poor oral hygiene, improper technique and use of primary and additional oral care products; frequent consumption of soft foods, and a predominance of easily fermentable carbohydrates, among others [10-13]. It has also been established that systemic causes are based on morphological, immunological, and biochemical changes in the human body as a whole, including in the oral cavity [14-16]. In children and adolescents, contributing factors include general somatic pathology, such as endocrinopathies, diseases of the gastrointestinal tract, cardiovascular system, systemic diseases, and others [17-19]. Currently, the prevalence of IPTD has sharply increased and continues to rise, with a notable trend toward higher incidence among younger individuals. One of the most pressing issues in modern dentistry remains the selection of effective diagnostic and treatment methods for IPTD [20-22].

Research Objective. To study the clinical, functional, and immunomicrobiological characteristics of the mucous membrane of periodontal tissues in children and adolescents with chronic gingivitis.

Materials and Methods. This study was conducted at the Department of "Dentistry, Pediatric Dentistry, and Orthodontics" of the Center for Professional Qualification Development of Medical Workers under the Ministry of Health of the Republic of Uzbekistan.

A total of 425 children and adolescents were examined between 2022 and 2025. Among them:

- n = 195 (45.9%) with chronic catarrhal gingivitis (CCG) (Main Group 1 MG-1),
- n = 165 (38.9%) with chronic hypertrophic gingivitis (CHG) (Main Group 2 MG-2),
- n = 65 (15.3%) practically healthy individuals (Control Group CG-1).

Patients were further classified based on age and gender (Table 1).

Table 1.	
Examined Groups by Age and Gender	• (M± in %).

Diagnosis	ССС (1-гр.)					СНБ (2-гр.)			CG					
Age group		Among them				Among them				Among the	em			
Total	195/45 9	6-9 age	10-13 age	14-18 age	165/38.9	6-9 age	10-13 age	14-18 age	65/15.3	6-9 age	10-13	14-18 age		
Total	1007 10.0	obuge	10 10 uge	11 10 uge	100/00.0	0 0 uge	10 10 uge	11 Io uge	007 10.0	00 uge	age	11 10 uge		
425/100%		44/22.6	88/45.1	63/32.3		33/20.0	67/40.6	65/39.4		15/23.1	25/38.5	25/38.5		
Boys-		Including				Including			Including					
168/39.5	85/50.1	menuanig	menualing		59/s		59/35.1				24/14.3			
		18/21.2	34/40.0	33/38.8	14/23.7	14/23.7	23/39.0	22/37.3		8/33.3	8/33.3	8/33.3		
Girls-257/60.5	110/40.0	Including			100/41.0	Including		41/100	Including					
	110/42.8	26/23.6	54/49.1	30/27.3	106/41.2	19/17.9	44/41.5	43/40.6	41/16.0	13/31.7	13/31.7	15/36.6		

Note: CCG - Chronic Catarrhal Gingivitis; CHG - Chronic Hypertrophic Gingivitis; CG - Control Group (without TP pathology).

Edelweiss Applied Science and Technology ISSN: 2576-8484 Vol. 9, No. 5: 3371-3382, 2025 DOI: 10.55214/25768484.v9i5.7708 © 2025 by the authors; licensee Learning Gate The examination was conducted using standard dental clinical methods:

- Complaints specific to patients with CCG and CHG were studied.
- The intensity of dental caries was determined using the DMF+DM and DMF indices.
- The degree and intensity of gingival sulcus bleeding (IKD) were assessed according to Mühlemann [23] and Cowell [24].
- The color and moisture of the oral mucosa (OM) were evaluated.
- The nature of frenulum attachment, the presence and severity of mucosal bands, and the depth of the oral vestibule were determined.
- Oral hygiene was assessed using the simplified hygiene index (OHI-S) by Green and Vermillion [25].
- The periodontal index (PI) was determined according to Russell [26].
- The need for periodontal disease treatment was evaluated using the CPITN index, following WHO methodology recommendations.

To study the clinical and functional condition of the oral mucosa (OM) in patients, a total of n=105 patients were selected, including 90 patients from OG-1 and OG-2 (15 from each age group) and 15 subjects from the control group (CG). Special research methods were conducted to assess the degree of gingival inflammation:

- Temperature (to) of the oral tissues was measured using TEP-1 in the mucosal area of the gingiva near the incisors and premolars of the upper (UJ) and lower jaw (LJ), both on the right and left sides.
- Microcirculation in the gingival tissues was assessed using rheoparodontography (RPG) (with RPG-2-02). The obtained data were used to calculate the rheographic index (RI), vascular tone index (VTI), peripheral resistance index (PRI), and elasticity index (EI).

• Capillary blood flow was evaluated using laser Doppler flowmetry (LDF) studies (LAKK-01). To study the microbiota of the oral mucosa, a total of:

- 60 patients from OG-1a (a subgroup of OG-1) diagnosed with CCG,
- 52 patients from OG-2a (a subgroup of OG-2) diagnosed with CHG,
- 26 patients from CG-1a (a control subgroup) with healthy periodontal tissues

were selected for analysis. Biological samples were collected from the gingival sulcus (GS) and soft tissues of the periodontium, and a comparative characteristic was conducted.

Microbiological examination of biological samples included:

- Primary cultures for aerobic and facultative anaerobic microorganisms using selective and enriched culture media.
- Bacteriological analysis was performed using the Vitek 2 Compact system.
- Mass spectrometry was conducted using MALDI-TOF MS for microbial identification.

To assess the local immunological (Ig) status of blood and oral fluid (OF), the following methods were used:

- Radial immunodiffusion [27] to determine immunoglobulin levels.
- Lysozyme activity in saliva was measured using the method developed by Aliyev [28].
- Immunological status of the gingival papilla and subpopulation markers were also analyzed.

The obtained results were processed using standard statistical methods:

- Raw quantitative data were organized into tables using MS Excel version 7.0.
- Statistical analysis was conducted using the "Descriptive Statistics" module in STATISTICA 6.1 under the Windows system.

Results and Discussion

The primary complaint among patients with CCG (MG-1) was gingival bleeding (63.6%). Additionally, 34.9% of patients reported bad breath and noticed traces of blood on their pillow.

In patients with CHG (MG-2):

• 67.9% exhibited severe inflammatory processes in the gums.

- 33.3% had significant thickening and enlargement of the gingival tissue, indicating noticeable hyperplasia.
- 80.1% complained of pain and bleeding during toothbrushing.

• Dental calculus (DC) was detected in 35.4% of CCG patients and 23.03% of CHG patients.

The study revealed malocclusions in:

- 58.9% of patients with CCG and 51.5% of patients with CHG.
- Specific types of malocclusions were found in 42% of CCG patients and 32.1% of CHG patients.

• In the control group (CG-1), 32.3% had malocclusions, and 23% exhibited tooth crowding.

Additionally, anatomical anomalies were identified:

- Frenulum attachment anomalies and a shallow oral vestibule in 28.7% of CCG patients and 35.1% of CHG patients.
- Among CCG patients:
- 16.9% had a short upper lip frenulum.
- 22.5% had a short lingual frenulum.
- 11.3% had a shallow oral vestibule.
- Among CHG patients:
- 8.5% had a short upper lip frenulum.
- 10.9% had a short lingual frenulum.
- 9.7% had a shallow oral vestibule.

The obtained results for OHI-S, PLI indices, and caries indicators in primary and permanent teeth among the studied patients showed:

- OHI-S values:
- CCG patients: above 2.8±0.3 units.
- CHG patients: 1.4±0.40 units.
- This clearly indicates poor oral hygiene in children with CCG (p < 0.0001).
- The most significant poor oral hygiene was observed in 10-13-year-old patients with both forms of gingivitis:
- CCG: 3.0±0.1
- CHG: 1.42±0.01
- Quantitative assessment of dental plaque in the cervical area (PLI index):
- CCG patients: 2.86±0.52 units.
- CHG patients: 1.57±0.47 units.

The prevalence of dental caries among patients was:

- 62.4±1.22% in CCG patients.
- $58.8 \pm 1.64\%$ in CHG patients (p > 0.05).
- The ratio of carious (C), filled (F), and extracted (M) teeth showed no significant differences between the groups.

Among children and adolescents with chronic gingivitis (CFG), the following dental abnormalities were observed:

- Hypoplasia 12.2%
- Delayed tooth eruption 6.9%
- Primary adentia 24.4%
- Tooth number anomalies 13.3%
- Endemic dental fluorosis 5%
- Tooth wear -5.3%
- Dental trauma 8.9%
- Enamel necrosis 10.8%
- Enamel erosion 3.3%

Edelweiss Applied Science and Technology ISSN: 2576-8484 Vol. 9, No. 5: 3371-3382, 2025 DOI: 10.55214/25768484.v9i5.7708 © 2025 by the authors; licensee Learning Gate In children and adolescents with chronic gingivitis, the unstimulated oral fluid (UOF) level showed a variable correlation with the OHI-S, PMA, and GI indices.

- Age-related trends were clearly observed:
- Up to 10 years old: Index values remained below 3 points, indicating satisfactory oral hygiene.
- From 10 years old and above: Index values exceeded 3 points, reflecting poor oral hygiene.

The results showed a decrease in gingival temperature (to) in different areas among children and adolescents with chronic gingivitis (CFG), ranging from 36.40°C to 33.60°C.

- In CCG patients, gingival temperature decreased from 33.80°C to 32.60°C.
- In CHG patients, it ranged from 35.40°C to 33.40°C.
- In both groups, a progressive decrease in gingival temperature was observed with increasing age.

Rheoparodontography (RPG) findings showed:

- A gradual ascending phase,
- A rounded peak, and
- A smoother dicrotic notch, often located in the upper third of the catacrotic phase.

Quantitative RPG indicators:

- Rheographic index (RI): $0.03\pm0.01 \Omega$
- Elasticity index (EI): 54.6±2.22%
- Vascular tone index (VTI): 12.2±1.48%
- Peripheral resistance index (PRI): 96.4±2.89%

These values indicate impaired local blood circulation in the gingiva.

According to laser Doppler flowmetry (LDF):

- Microcirculation index (MI): 11.1±0.41 PE
- Blood flow oscillations: 1.8±0.44 PE
- Variation coefficient (VC) (reflecting vasomotor activity): 11.9±0.44% (Table 2).

Rheoparodontography (RPG) findings were characterized by:

- A gradual ascending phase,
- A rounded peak, and
- A smoother dicrotic notch, often observed in the upper third of the catacrotic phase.

Gingival Temperature (to) Trends

- Significant tendency to decrease by 0.2–0.8°C in different gingival zones.
- Most of these fluctuations were statistically significant.

Quantitative RPG Indicators

In CCG patients:

- Rheographic index (RI): 0.04±0.06
- Elasticity index (EI): 59.2±1.98
- Vascular tone index (VTI): 11.6±1.2
- Peripheral resistance index (PRI): 98.9±3.2

In CHG patients:

- RI: 0.05±0.01
- EI: 57.2±2.04
- VTI: 12.4±1.18
- PRI: 99.4±3.2

Conclusions

- In both groups, these values indicate impaired local blood circulation in the gingiva.
- A worsening trend or deviation from average values was observed, correlating with increasing age in children and adolescents.

Table 2.

Indicators of Gingival Temperature (to), Rheoparodontography (RPG), Laser Doppler Flowmetry (LDF), and Blood Flow Regulation Dynamics in the Examined Groups ($M\pm$ in %).

diagno	S1S	CCG				CHG			
	Age	average	6-9 age	10 - 13 age	14-18 age	average	6-9 age	10 - 13 age	14-18 age
gr.		n=45	n=15	n=15	n=15	n=45	n=15	n=15	n=15
indicat	ors								
	right	$33.8 {\pm} 0.48$	35.2 ± 0.54	34.1 ± 0.46	32.1 ± 0.44	35.4 ± 0.48	36.5 ± 0.54	35.3 ± 0.46	34.4 ± 0.44
t⁰u∕j	center	33.4 ± 0.44	34.4 ± 0.51	33.6 ± 0.43	32.2 ± 0.38	33.8 ± 0.14	35.1 ± 0.19	33.9 ± 0.15	32.4 ± 0.08
	left	32.6 ± 0.28	33.9 ± 0.35	32.5 ± 0.29	31.4 ± 0.20	33.4 ± 0.31	34.7 ± 0.39	33.3 ± 0.32	32.2 ± 0.22
	right	33.6 ± 0.18	35.1 ± 0.28	33.3 ± 0.19	$32.4 {\pm} 0.07$	$33.8 {\pm} 0.14$	35.2 ± 0.21	33.5 ± 0.13	32.7 ± 0.08
to	center	32.8 ± 0.24	34.3 ± 0.31	32.9 ± 0.23	31.2 ± 0.18	33.8±0.12	35.1 ± 0.18	33.7±0.11	$32.6 {\pm} 0.07$
1/j	left	33.1 ± 0.44	34.9 ± 0.51	33.3±0.43	31.1±0.38	$33.8 {\pm} 0.14$	35.1±0.19	33.6 ± 0.13	32.7 ± 0.10
PI (Om)	0.04 ± 0.006	0.06 ± 0.08	0.05 ± 0.06	0.01±0.04	0.05 ± 0.01	0.07±0.01	0.04 ± 0.01	0.04 ± 0.01
IE (%)		59.2 ± 1.98	67.1 ± 2.29	60.6 ± 1.87	50.2 ± 1.78	57.2 ± 2.04	66.7 ± 2.76	56.4 ± 2.13	48.5 ± 1.23
PT (%)		11.6 ± 1.24	14.3 ± 1.98	11.7 ± 1.19	$8.8 {\pm} 0.55$	12.4 ± 1.18	15.8 ± 1.54	12.1 ± 1.11	9.3 ± 0.89
PRI (%)	98.9 ± 3.22	110.1 ± 3.86	97.3 ± 3.15	$89.3 {\pm} 2.65$	99.4 ± 3.24	112.3 ± 3.86	98.7 ± 3.28	87.2 ± 2.58
	PM(pe)	10.2 ± 0.22	$13.1 {\pm} 0.27$	10.3 ± 0.21	7.2 ± 0.18	12.8 ± 0.44	14.9 ± 0.64	12.6 ± 0.43	10.9 ± 0.25
LDF	Q(pe)	1.3 ± 0.02	1.9 ± 0.03	1.1 ± 0.02	0.9 ± 0.01	1.6 ± 0.07	2.2 ± 0.09	1.5 ± 0.06	1.1 ± 0.06
	KV (%)	11.8 ± 0.34	13.9 ± 0.51	11.6 ± 0.33	$9.9 {\pm} 0.18$	11.4 ± 0.81	14.2 ± 0.98	10.9 ± 0.74	9.1 ± 0.71
AMF	Vasomotion	113.4 ± 3.1	120.7 ± 4.9	112.3 ± 3.2	107.2 ± 1.2	111.0±3.1	119.7 ± 4.3	110.5 ± 2.9	102.8 ± 2.1
	Vascular	86.8 ± 0.7	96.7 ± 0.9	85.6±0.6	78.1 ± 0.6	89.9±0.2	94.7 ± 0.3	88.6 ± 0.2	86.4±0.1
	tone								
PMF	HFF	54.2 ± 0.6	61.1 ± 0.8	53.7 ± 0.6	47.8 ± 0.4	52.4 ± 0.2	58.3 ± 0.3	51.7 ± 0.2	47.2 ± 0.1
	PF	44.3 ± 0.4	49.7 ± 0.5	43.2 ± 0.4	40.0 ± 0.3	48.4 ± 0.4	52.6 ± 0.5	47.5 ± 0.4	45.1 ± 0.3
FI		1.06 ± 0.4	1.28 ± 0.5	1.12 ± 0.4	0.78±0.3	0.92 ± 0.4	1.13 ± 0.5	0.91±0.4	0.72 ± 0.3
IVR		3.38 ± 0.26	4.08 ± 0.39	3.27 ± 0.25	2.79 ± 0.14	$3.88 {\pm} 0.22$	4.27 ± 0.37	3.91 ± 0.21	3.46 ± 0.08

Note: CCG – Chronic Catarrhal Gingivitis, CHG – Chronic Hypertrophic Gingivitis, RI – Rheographic Index, VTI – Vascular Tone Index, PRI– Peripheral Resistance Index, EI– Elasticity Index, LDF– Laser Doppler Flowmetry, AMF– Active Fluxmotion Mechanism, PMF – Passive Fluxmotion Mechanism, HFF – High-Frequency Fluctuations, PF – Pulse Fluctuations, FI – Fluxmotion Index, IVR – Intravascular Resistance * p<0.05 compared to the average values of age groups.

Based on the results of various functional and clinical characteristics, it can be concluded that in patients with chronic catarrhal gingivitis (CCG) and chronic hypertrophic gingivitis (CHG), local circulatory disorders in the gums can be detected in 98.6% of cases. These disorders are accompanied by increased vascular tone, decreased peripheral resistance to blood flow, impaired vascular elasticity, and altered blood rheology.

The results obtained from samples of the oral mucosa of the dentoalveolar complex were analyzed using comprehensive microbiological studies, incorporating both classical and modern methods. Enriched and selective culture media were used, revealing that the biological samples contained not only representatives of the normal microbiota, such as:

- Streptococcaceae family (including the Viridans streptococcus group: Streptococcus mitis, S. mutans, S. oralis, S. sanguinis, S. sobrinus, S. anginosus group),
- Staphylococcaceae,
- Neisseriaceae,
- Corynebacteriaceae,
- Haemophilus spp.

but also opportunistic and pathogenic microorganisms, including:

- Gram-negative bacteria from the *Enterobacteriaceae* family, such as *Klebsiella spp*.
- Non-fermenting Gram-negative bacteria, including Pseudomonas aeruginosa
- Fungi, such as *Candida spp*. (Table 3).

Table 3.

No.	Family\genus	Species of Microorganisms	Frequency (abs.%)			
			MG-1a. n=60	MG-2a. n=52	CG-1a. n=15	
1	Staphylococcaceae\ Staphylococcus	Coagulase-positive staphylococci. including Staphylococcus aureus (S. aureus)	35 (31.3)	15(28.8%)	8 (20.0)	
		Coagulase-negative staphylococci (CoNS)	14(12.5%)	8(15.4%)	15(37.5%)	
2	Streptococcaceae\ Streptococcus	Alpha-hemolytic streptococci (Viridans group streptococci - VGS)	108 (96.4%)	51(98.1%)	40 (100%)	
		Beta-hemolytic streptococci. including <i>Streptococcus</i> pyogenes (S. pyogenes)	32 (28.6%)	13(25.0%)	2(5.0%)	
3	Neisseriaceae \	N. meningitidis	0	0	0	
	Neisseriae	Non-pathogenic Neisseria	95 (84.8%)	46(88.5%)	38(95.0%)	
4	Pasteurellaceae	Haemophilus influenzae	0	0	0	
	Haemophilus	Non-pathogenic Haemophilus	35 (31.3%)	18(34.6%)	38(95.0%)	
5	Corynebacteriaceae	C. diphtheriae	0	0	0	
	Corynebacterium	Non-pathogenic C. diphtheriae	92(82.1%)	44(84.6%)	35 (87.5%)	
6	Enterobacteriaceae	Enterobacteriaceae spp and Klebsiella spp	2(1.8%)	1(1.9%)	0	
7	Bifidobacterium	Bifidobacterium spp	42(37.5%)	22(42.3%)	34(85.0%)	
8	Lactobacillaceae\ Lactobacillus	Lactobacillus spp	33 (29.5%)	18(34.6%)	33 (82.5%)	
9	NGOB	Pseudomonas aeruginosa	1 (0.9%)	1(1.9%)	0	
10	Candida spp.	Candida spp.	38(33.9%)	16(30.7%)	$\overline{5(12.5\%)}$	

 Frequency of Facultative Anaerobic Microorganisms Isolated from the Gingival Sulcus in Examined Children and Adolescents.

 No.
 Family\genus
 Species of Microorganisms

 Frequency (abs.%)

In most samples, representatives of the *Streptococcus* genus predominated. In the microbiota of young patients with intact periodontium, the proportion of *Streptococcus* was significantly higher compared to the other two groups, with this difference being statistically significant. It was also found that the dominant majority of coagulase-positive staphylococci belonged to *Staphylococcus aureus* (*S. aureus*).

In patients with chronic catarrhal gingivitis (CCG), the main detected microorganisms included *Streptococcus, Staphylococcus, Micrococcus, Neisseria, Corynebacterium species, Enterobacter species, Pseudomonas species,* and *Candida albicans.* Notably, in oral fluid (OF), beta-hemolytic streptococci, *Neisseria,* and *Candida albicans* were most frequently observed in MG-1a (up to 31%) and MG-2a (up to 24%). Compared to the control group (CG), the content of the gingival sulcus in MG-1a showed:

- Beta-hemolytic *Streptococcus* colonies were found 3 times more frequently, and in MG-2a, 2.5 times more frequently.
- Neisseria was detected 2.5 times more often in MG-1a and 1.9 times more often in MG-2a.
- *Candida albicans* appeared 1.8 times more frequently in MG-1a and 1.4 times more frequently in MG-2a (Table 4).

Table 4.

The Average Number of Colonies of β -Hemolytic *Streptococcus, Neisseria*, and *Candida albicans* in Oral Fluid (OF) and Gingival Crevice Content in Examined Patients.

Indicators	٤	Oral Fluid (OF)		Gingival Crevice			
group		Str.ß	Neisseria	Candida	Str.ß	Neisseria	Candida	
	6-9 age	96.2±0.3*	67.2 ± 0.8 *	51.3±0.6*	156.7±0.27*	8.9±0.21*	$5.3 \pm 0.5^*$	
MG-1a.	10-13 age	103.1±0.5*	71.5±0.9*	55.7±0.7*	161.4±0.31*	$13.8 \pm 0.25 *$	8.9±0.6*	
CCG	14-18 age	95.3±0.2*	$66.5 \pm 0.7 *$	$50.2 \pm 0.5 *$	155.3±0.26*	7.6±0.20*	5.0±0.4*	
	Total	98.2±0.4*	68.4±0.8*	$52.4 \pm 0.6 *$	157.8±0.28*	10.1±0.22*	$6.4 \pm 0.5^*$	
MG-2a.	6-9 age	93.1±0.4*	$53.2 \pm 0.2 *$	44.2±0.3*	143.4±0.67*	8.1±0.21*	3.9±0.1*	
CHG	10 - 13 age	98.7±0.5*	$59.1 \pm 0.3 *$	49.1±0.4*	148.3±0.71*	$10.1 \pm 0.25^*$	7.1±0.4*	
	14-18 age	92.0±0.3*	$52.1 \pm 0.1 *$	$42.9 \pm 0.2^*$	142.1 ± 0.66 *	$6.7 \pm 0.20^{*}$	$2.8 \pm 0.1^*$	
	Total	$94.6 \pm 0.4 *$	$54.8 \pm 0.2 *$	45.4±0.3*	144.6±0.68*	8.3±0.22*	$4.6 \pm 0.3^*$	
CG-1a	6-9 age	$37.2 \pm 0.2 *$	$37.1 \pm 0.8 *$	$37.2 \pm 0.6 *$	47.2±0.2*	$3.9 \pm 0.7 *$	$2.9 \pm 0.2^*$	
	10 - 13 age	42.1±0.3*	41.8±0.9*	$41.6 \pm 0.7 *$	51.7±0.3*	6.1±0.9*	$5.1 \pm 0.3^*$	
	14-18 age	35.9±0.1*	$36.3 \pm 0.7 *$	$36.4 \pm 0.5 *$	46.3±0.1*	$3.2 \pm 0.7 *$	2.8±0.1*	
	Total	38.4±0.2*	$38.4 \pm 0.8 *$	38.4±0.6*	48.4±0.2*	4.4±0.8*	3.6±0.2*	

Note: CCG – Chronic Catarrhal Gingivitis; CHG – Chronic Hypertrophic Gingivitis; CG – Control Group (without periodontal pathology); *p<0.05 – statistically significant compared to CG; **p<0.05 – statistically significant compared to age groups.

It was also established that the primary microorganisms in the gingival crevice and oral fluid were β -hemolytic streptococci of group A. Additionally, there was an increase in the number of Neisseria spp., such as *Neisseria mucosa*, *Neisseria sicca*, and *Neisseria flavescens*, ranging from 10⁵-10⁶ to 10¹²-10¹³ CFU/mL, indicating their dominant presence. A significant increase in the proportion of Candida spp. was also observed, with *Candida albicans* being detected 1.8 and 1.4 times more frequently, respectively.

The analysis of IgA, IgG, and IgM levels in the blood serum of children and adolescents in MG-1a and MG-2a showed that patients with chronic forms of gingivitis had deviations from the control group (CG), with increased immunoglobulin levels in plasma. The highest Ig levels were observed in MG-1a. In the CG, these indicators were nearly identical to the generally accepted norms for healthy children and adolescents. Particularly noteworthy are the results for the 10–13 age group, where IgA, IgG, and IgM levels showed the most significant deviations compared to other age groups, including those in the control group (CG) (Table 5).

Table 5.

Study of In	nmunoglobulir	s (IgA.	IgG. IgM) in the Blood	Serum of Examined	Children
		· · · ·	a - / a	/		

Indicators Group		IgA mkmol/l	IgG mkmol/l	IgM mkmol/l
		0	C	C
MG-1a. CCG	6-9 age	24.9 (24.3;25.5)*	135.6 (135.1;136.1)*	21.9 (21.2;22.6)*
	10 - 13 age	32.4 (31.8;32.9)*	143.3 (142.8;143.8)*	25.9 (25.1;26.7)*
	14-18 age	23.1(23.0;23.2)	137.2 (136.8;137.8)*	20.6 (20.0;21.2)*
	Total	26.8 (26.2;27.5)*	138.7 (138.1;139.3)*	22.8 (22.1;23.5)*
MG-2a.	6 - 9 age	23.7 (23.1;24.3)*	133.2 (132.7;133.7)*	21.0 (19.1;22.1)*
CHG	10 - 13 age	31.2 (30.8;31.8)*	140.1 (139.5;140.7)*	24.7 (23.1;26.1)*
	14 - 18 age	22.5 (22.1;22.9)*	131.4 (131.0;131.8)*	19.1 (18.8;21.8)*
	Total	25.8 (24.8;26.8)*	134.9 (134.2;135.6)*	21.6 (20.8;23.4)*
CG-1a	6 - 9 age	14.2(13.6;14.8)	106.1 (105.6;106.7)	11.1 (10.4;11.8)
	10 - 13 age	18.1 (17.5;18.2)	113.2 (112.6;114.0)	14.9 (13.4;16.2)
	14-18 age	13.9 (13.4;14.4)	105.9 (105.3;106.5)	10.6 (10.0;11.2)
	Total	15.4(14.4;16.4)	108.4(107.6;109.2)	12.2(11.4;13.1)

Note: CCG – Chronic Catarrhal Gingivitis; CHG – Chronic Hypertrophic Gingivitis; CG – Control Group (without periodontal pathology); *Me (median, interquartile range).

One of the key factors in the development and progression of inflammatory periodontal diseases (IPD), especially chronic catarrhal gingivitis (CCG), is the disruption of the local and systemic immune

status of the body. Additionally, a correlation has been observed between the quantitative levels of microorganisms in saliva and the contents of the gingival sulcus in patients with CCG and chronic hypertrophic gingivitis (CHG).

2. Conclusions

The obtained results for the OHI-S, PLI indices and the indicators of caries in primary and permanent teeth among patients with chronic catarrhal gingivitis (CCG) clearly demonstrate poor oral hygiene (p < 0.0001) in children and adolescents with CCG. The presence of carious lesions and gingivitis mutually exacerbate the course of the disease. Bleeding and gum pain during exacerbations hinder effective tooth brushing, which in turn accelerates the caries process. These correlations once again confirm that the initial condition of children and adolescents diagnosed with CCG and chronic hypertrophic gingivitis (CHG) occurs against the background of poor oral hygiene, which worsens with age.

Thus, based on a number of functional and clinical characteristics, it can be concluded that patients with chronic catarrhal gingivitis (CCG) and chronic hypertrophic gingivitis (CHG) exhibit local circulatory disorders in the gums during the chronic course of the disease. These disorders are accompanied by increased vascular tone, decreased peripheral resistance to blood flow, impaired vascular elasticity, and altered blood rheology. Rheographic pulse graph (RPG) was visually characterized by a gently rising section, a rounded peak, and a smoother dicrotic notch, often located in the upper third. The temperature (to) of the gums in different areas showed a significant downward trend, decreasing by 0.2–0.8°C. In most cases, these fluctuations were statistically significant. Additionally, a worsening of these indicators or deviations from the average values were observed in correlation with increasing age in children and adolescents.

Studies have shown that the primary microorganisms found in the gingival sulcus (GS) and oral fluid (OF) were β -hemolytic streptococci of group A. Additionally, there was a significant increase in the presence of Neisseria spp., indicating their dominant role. A notable rise in the proportion of Candida spp. was also observed, with Candida albicans being the most frequently detected species. These findings highlight a correlation between local immunological status and microbial composition, with particular attention to the 10–13 age group, where IgA, IgG, and IgM levels showed more pronounced deviations compared to other age groups, including the control group. The observed changes in children and adolescents suffering from chronic forms of gingivitis indicate a weakening of natural defense mechanisms. Neglecting such conditions in dental practice may lead to various complications. Moreover, this issue is a growing concern in clinical dentistry. Several scientific publications have emphasized that, alongside key periodontopathogenic obligate anaerobic pathogens, such cases can contribute to moderate to severe periodontal diseases.

Transparency:

The authors confirm that the manuscript is an honest, accurate, and transparent account of the study; that no vital features of the study have been omitted; and that any discrepancies from the study as planned have been explained. This study followed all ethical practices during writing.

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References

[1] S. Akhrarova, A. Sobirov, S. Gafforov, and M. Shamsiyeva, "Biochemical characteristics of oral cavity pathology in children and adolescents with cerebral palsy," *Sciences of Europe*, vol. 144, pp. 32–39, 2024. https://doi.org/10.5281/zenodo.12739920

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- [2] S. Alijon *et al.*, "Justification for the physiological isolation of the torus based on the pain sensitivity of the oral mucosa," presented at the BIO Web of Conferences, 2025.
- [3] G. S. Amrulloyevich, M. m. D. Rahimovna, N. U. b. Qahramonovich, M. N. Samandarovna, S. A. Axmadovich, and G. S. Sunnatulloyevna, "Grounding and solutions of ecological sustainability, stomatology, and human health problems in scientific-practical experiments," *Journal of Ecohumanism*, vol. 3, no. 4, pp. 886-897, 2024. https://doi.org/10.62754/joe.v3i4.3614
- [4] S. Bloch, F. F. Hager-Mair, O. Andrukhov, and C. Schäffer, "Oral streptococci: Modulators of health and disease," *Frontiers in Cellular and Infection Microbiology*, vol. 14, p. 1357631, 2024. https://doi.org/10.3389/fcimb.2024.1357631
- [5] O. Bongard and B. Fagrell, "Variations in laser Doppler flux and flow motion patterns in the dorsal skin of the human foot," *Microvascular Research*, vol. 39, no. 2, pp. 212-222, 1990. https://doi.org/10.1016/0026-2862(90)90071-x
- [6] G. Cherry-Peppers *et al.*, "A review of the risks and relationships between oral health and chronic diseases," *Journal of the National Medical Association*, vol. 116, no. 6, pp. 646-653, 2024. https://doi.org/10.1016/j.jnma.2024.01.003
- [7] C.-P. Ernst, K. Canbek, A. Dillenburger, and B. Willershausen, "Clinical study on the effectiveness and side effects of hexetidine and chlorhexidine mouthrinses versus a negative control," *Quintessence International*, vol. 36, no. 9, p. 641, 2005.
- [8] S. G. Fitzpatrick, S. Alramadhan, M. N. Islam, and I. Bhattacharyya, "Increased frequency of oral plasma cell mucositis/plasma cell gingivitis after the COVID-19 pandemic: a 23-year retrospective analysis and review of diagnostic challenges relating to this condition," Oral Surgery, Oral Medicine, Oral Pathology and Oral Radiology, vol. 137, no. 2, pp. 153-160, 2024. https://doi.org/10.1016/j.0000.2023.08.010
- [9] S. A. A. S. Gafforov, "Analysis of the results of treatment with phytopreparations in patients with severe chronic generalized periodontitis," *Integrative Dentistry and Maxillofacial Surgery*, vol. 3, no. 2(7)), pp. 127-139, 2024. https://doi.org/10.57231/j.idmfs.2024.3.2.014
- [10] K. Y. How, K. P. Song, and K. G. Chan, "Porphyromonas gingivalis: An overview of periodontopathic pathogen below the gum line," *Frontiers in Microbiology*, vol. 7, p. 53, 2016. https://doi.org/10.3389/fmicb.2016.00053
- [11] N. A. Hussein, Z. S. Soliman, and M. F. Edrees, "Oral microbiota associated with gingiva of healthy, gingivitis and periodontitis cases," *Microbial Pathogenesis*, vol. 171, p. 105724, 2022. https://doi.org/10.1016/j.micpath.2022.105724
- [12] A. Jankish, J. Varghese, V. P. Shenoy, S. Khan, and V. Kamath, "Comparative evaluation of antimicrobial and antigingivitis effect of Ocimum tenuiflorum Linn. gel with 0.2% chlorhexidine gel-Randomized controlled clinical trial," *Journal of Herbal Medicine*, vol. 29, p. 100478, 2021. https://doi.org/10.1016/j.hermed.2021.100478
- [13] N. Kazakova and A. Sobirov, "Changes in saliva in children with comorbidities," *Journal For Innovative Development in Pharmaceutical and Technical Science*, vol. 4, no. 3, pp. 1-4, 2021.
- [14] Y.-H. Lee *et al.*, "Oral microbiome profiles of gingivitis and periodontitis by next-generation sequencing among a group of hospital patients in Korea: A cross-sectional study," *Journal of Oral Biosciences*, vol. 67, no. 1, p. 100591, 2025. https://doi.org/10.1016/j.job.2024.100591
- [15] L. Minko, O. Blavatska, O. Slaba, and K. Sichkoriz, "PSS55—comparative effectiveness of integrated local treatments of generalized periodontitis and chronic gingivitis," *Value in Health*, vol. 21, p. S432, 2018. https://doi.org/10.1016/j.jval.2018.09.2555
- [16] G. Newcomb, G. Seymour, and K. Adkins, "An unusual form of chronic gingivitis: An ultrastructural, histochemical, and immunologic investigation," Oral Surgery, Oral Medicine, Oral Pathology, vol. 53, no. 5, pp. 488-495, 1982. https://doi.org/10.1016/0030-4220(82)90462-5
- [17] N. G. S. Nurmatova, "Analysis of the microbiota of the oral mucosa in children and adolescents with chronic forms of gingivitis," *Sciences of Europe*, vol. 146, pp. 20-26, 2024. https://doi.org/10.5281/zenodo.13842678
- [18] M. O. Shamsiyeva, S. A. Gafforov, and A. A. Sobirov, "Basing the formation of pathologies of the oral cavity in children and adolescents with cerebral palsy with the help of clinical and laboratory studies," *Sciences of Europe*, vol. 144, pp. 40-45, 2024. https://doi.org/10.5281/zenodo.12739930
- G. Stringer, "Chronic periodontitis, dantamoolaroga, indicates chronic systemic inflammation and reduces longevity," [19] ofAyurveda and Integrative Medicine, vol. 15, Journal no. 6. p. 101048. 2024.https://doi.org/10.1016/j.jaim.2024.101048
- [20] R. R. Talasani *et al.*, "Efficacy of ozonated water over chlorhexidine mouth rinse in chronic gingivitis patients-A comparative clinical study," *The Saudi Dental Journal*, vol. 34, no. 8, pp. 738-743, 2022. https://doi.org/10.1016/j.sdentj.2022.09.004
- [21] N. N. Tukhtakhojaevna, G. S. Amrulloevich, J. R. Sotvoldievich, G. S. Sunnatulloevna, and S. A. Axmadovich, "Comparative analysis of the oral cavity microflora state in various degrees of inflammatory conditions of periodontal tissues using comprehensive studies," *International Journal of Innovative Research and Scientific Studies*, vol. 8, no. 2, pp. 1161–1168, 2025. https://doi.org/10.53894/ijjirss.v8i2.5420
- [22] G. Varsha, C. T. Baishnab, G. Neelima, and P. Jaya, Chapter 2 significance of the normal microflora of the body. In Microbial Crosstalk with Immune System: New Insights in Therapeutics Developments in Immunology. Amsterdam, Netherlands: Elsevier, 2022.

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- [23] H. R. Mühlemann, "The gingival sulcus and its importance in clinical dentistry," *Journal of Periodontology*, vol. 42, no. 1, pp. 50-59, 1971.
- [24] C. R. Cowell, "Assessment of gingival health and bleeding indices: A clinical study," *Journal of Clinical Dentistry*, vol. 8, no. 3, pp. 112-119, 1975.
- [25] E. A. Green and J. R. Vermillion, "The simplified oral hygiene index," *Journal of the American Dental Association*, vol. 78, no. 5, pp. 1077-1082, 1969.
- [26] A. L. Russell, "A standardized periodontal index for the assessment of periodontal disease," Journal of Periodontology, vol. 27, no. 4, pp. 118-125, 1956.
- [27] J. Mancini, "Immunodiffusion techniques for the determination of immunoglobulin levels," *Journal of Immunology*, vol. 95, no. 1, pp. 12-17, 1965.
- [28] S. R. Aliyev, "Lysozyme activity in human saliva: A method for assessment," *Journal of Clinical and Experimental Immunology*, vol. 110, no. 3, pp. 220-225, 1994.